

**PHYSICAL PARAMETERS OPTIMIZATION FOR ENHANCEMENT OF PRODIGIOSIN PRODUCTION BY USING *SERRATIA SPP.*****<sup>1</sup>Phatake Y. B. and <sup>2</sup>Dharmadhikari S.M.**<sup>1</sup>Department of Microbiology, Vidya Pratishthan School of Biotechnology, Baramati, Maharashtra.<sup>2</sup>Department of Microbiology, Government Institute of Science, Aurangabad.**\*Corresponding Author: Phatake Y. B.**

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**ABSTRACT**

The nutrients require for the growth of the microorganisms is mainly classified as macro nutrients like carbon, nitrogen etc. and micronutrients like iron, calcium etc. The macro nutrients seriously affect growth of the microorganism and yield of the secondary metabolite. Light play important role in the life of animal and plant but microorganisms also show morphological and behavioral changes in presence of different wavelength of the light. So it is necessary to optimize the production process with respect to growth condition and nutrient level to get maximum yield of the product. In this study effect of different physical parameters including light on prodigiosin production was determined and process were further optimized by using classical approach. The study reveal the effect of pH of the medium, time of incubation, temperature of incubation, speed of agitation and different wavelength of light on the red pigment prodigiosin production and on the yield of biomass. As well as stability of produced pigment to the pH. Maximum yield of pigment was obtained in unit/ cell when producer organisms were inoculated in peanut broth of pH 7, incubated at 100rpm in 28°C temperature for 72hrs. In this study effect of spectrum of light on pigment and biomass production was also determined. Maximum pigment production in unit/ cell was observed when inoculated broth were subjected to white light followed by yellow light and blue light. The incubation in red and green light reduces the yield of pigment but they did not completely inhibit the synthesis of it. Increased pigment production was observed in total white light as compared to the flasks completely exposed to darkness. The visual comparison of the extracted pigment exhibited varied shades of color to different wavelengths of light. Broth exposed to white light incubation yielded a deep red colored pigment whereas yellow light incubation yielded a light red to orange colored pigment. The yield of biomass also changes according to pigment yield, white, blue and yellow light give high yield of biomass, whereas red light and green light reduced the cell growth. The stability of the pigment to pH was also determined. At extremely acidic pH, the red colour pigment appeared as pink (1 to 5). While in highly basic condition, the pigment was almost decolorized (9 to 13).

**KEYWORDS:** Prodigiosin, light spectrum, *Serratia spp.*, Biomass production.**INTRODUCTION**

From old days humans have traditionally preferred natural sources to add colors to food, clothing, cosmetics and medicinal product. Microbes have advantages of versatility and productivity over higher forms of life in the industrial-scale production of natural pigments and dyes. Microbial pigments have two meaningful advantages over artificial and inorganic colors. One relates to fermentation, which is an inherently faster and more productive production compared to any other chemical process. The other enduring strength of microbes is their relatively large and easily manipulated strands of genes (Canizares, V. R. O. *et al.*, 1998).

Nearly for all living organisms crucial environmental signals for regulating development and physiological

process are light, temperature, pH, humidity etc. Consequently, the capacity of organism to sense light signal and respond to it is extensive present in kingdoms, from archaea and fungi to humans (Arden, G. B. *et al.*, 1996 and Babitha, S. *et al.*, 2008). Several studies on the factors influencing the formation of the red pigment prodigiosin by *Serratia spp.* have been made. In general, such studies have included the influence of carbon source, nitrogen source, pH of the medium, time of incubation etc.

Light has a crucial influence on microorganisms due to its capability of inducing morphological and behavioral changes (Casas-Flores, *et al.*, 2006 and Soumya, K. *et al.*, 2014). The effect of different wavelength of light on pigment production in fungi was

well studied. Researcher had found that production of colour compounds in fungi significantly affect by intensity of light and its wavelength.

The researcher has found that light is also a very important factor which affects production of pigment and biomass not only in bacteria but also in Actinomycetes (Pearce, G. W. and Streeter, I. R. 1931). The capacity to sense and respond to light is wide spread in animals, plants, fungi and bacteria. Electrical signals have been detected in response to light excitation of the bacteria *Serratia marcescens*. These signals are related to the wavelength and intensity of the stimulus and the growth stage of the bacteria. (Babitha, S. et. al., 2008).

In the last decade, there has been a great increase in utilization of natural pigments in the food industries, replacing the implementation of artificial coloring agents, in order to provide a healthier food for consumers. However, in comparison with the natural coloring agents, the artificial coloring agents show greater resistance and stability when exposed to oxidation, changes in temperature, pH and other factors (Francis, 1989; Hong and Wrolstad 1990; She, *et al.*, 1992; Fabre, *et al.*, 1993, Laleh, *et al.*, 2006).

In this study effect of pH of the medium, time of incubation, temperature of incubation, speed of agitation and different wavelength of light on the red pigment production and on the biomass yield of producer organism were studied. As well as stability of produced pigment to the pH were also determined by exposing crude pigment to various pH.

## MATERIAL AND METHOD

For the present study bacterial strains of *Serratia marcescens* (YP 2, YP 10, YP 14 and YP16) previously isolated from soil and water samples and characterized by using Bergey's Manual of Determinative Bacteriology (Holt, *et al.*, 1989) and Benson's Microbiological Applications, Laboratory Manual in General Microbiology (Brown, 2007), (Nageswaran, N. *et al.*, 2014) was used (Phatake, Y. B. *et al.*, 2015). Strains were lyophilized and also maintained on nutrient agar slant at 4°C and stored at microbial culture collection unit of Vidya Pratishthan. School of Biotechnology, Baramati, Maharashtra, India. Inoculum was prepared in peanut broth (g/L): peanut powder 25, pH 7 by transferring a loop of the bacteria from stock culture and incubated at 37°C, for 24 hr.

### Optimization of Production Process

In order to determine the optimum condition for production of prodigiosin, selected strains of *Serratia marcescens* were grown in Peanut broth. For optimization of the pH, bacterium were inoculated in peanut broth having different pH viz. 5, 5.5, 6, 6.5, 7, 7.5, 8, 8.5, and 9, for optimization of time of incubation organism was incubated for different time interval viz. 24, 48, 72 and 96 h. and then for optimization of

temperatures organism were incubated in temperatures Viz. 25°C, 28°C, 31°C, 34°C and 37°C for 72h. To check the effect of agitation on the yield of prodigiosin all four isolates were inoculated in the peanut broth and incubated at 28°C for 72 hours at different rpm viz. 00, 25, 50, 75, 100, 125.

All the experiments were carried out in triplicates and mean yield and standard error was calculated for each observation. Pigment yield in unit /Cell was calculated by using following formula (Mekhael R. and Yousif S.Y. 2009).

$$\text{Prodigiosin Unit/cell} = \frac{[\text{OD. 499} - (1.381 \times \text{OD 620})] \times 1000}{\text{OD 620}}$$

OD 499: Pigment absorbance.

OD 620: Bacterial cell absorbance.

1.381: Constant

### Effect of light spectrum on prodigiosin production.

On the basis of optimization studies, single potent pigment producing strain of *serratia* (YP 2) was cultured in 100 ml of Peanut Broth, to study the effect of different wavelengths of light on biomass and pigment production. For this experiment, instead of using different wavelengths of light, different colored gelatin papers were used. Because colored paper allow only selected wavelength of light to pass through it, whereas it filters out all other wavelengths. The flasks were wrapped in gelatin papers of red, blue, green and yellow colors. One flask was covered with black art paper to completely cut off the light and one flask was kept as control which was exposed to white light. All the flasks (Fig.No.2) were placed at equidistant (15cm) from the illuminated light source (Philips CFL 25 watts) after inoculation (0.1 ml) of selected strain of bacteria (Soumya, K. *et al.*, 2014).

Experiment was carried out for five days (120 hrs). After regular interval of time (24 h), sample was collected from each flask aseptically and analyzed for pigment production, biomass production, pH change and colony morphology. For the extraction and quantification of the prodigiosin, broth containing pigment was mixed with equal volume of methanol and kept the mixture in rotary shaker for 20 to 30 min. Then this mixture was pour into the centrifuge tube and shaken vigorously using a vortex mixture. Then centrifuge at 10000 rpm for 10 min. The supernatant was collected and filtered through Wattman filter paper number 1 (0.2µm). The filtrate was concentrated using a rotary evaporator and later extracted with 3.0 M chloroform. The chloroform phase was collected and reconcentrated using rotary evaporator to obtain crude powder. Yield in Unit/Cell was calculated by spectroscopic analysis (Davaraj, *et al.*, 2009). Concentration of the extracted Prodigiosin was estimated in unit/cell.

**Effect of light spectrum on biomass.**

After regular time interval cell mass was separated from the broth by centrifugation and filtration. The separated cell mass was washed thrice with distilled water and concentrated in hot air oven at 80°C and the concentrated biomass was weighed in a pre-weighed beaker to estimate dry weight.

**Stability of pigments to different pH.**

The extracted red colour pigment prodigiosin was tested for its stability to pH ranging from extremely acidic (1) to highly basic (13). For this, pigment dissolve in methanol was distributed in seven tubes and pH of each vial was adjusted with the help of 0.1N HCL and NaOH. The observation in the form of change in colour and change in lambda max value was recorded.

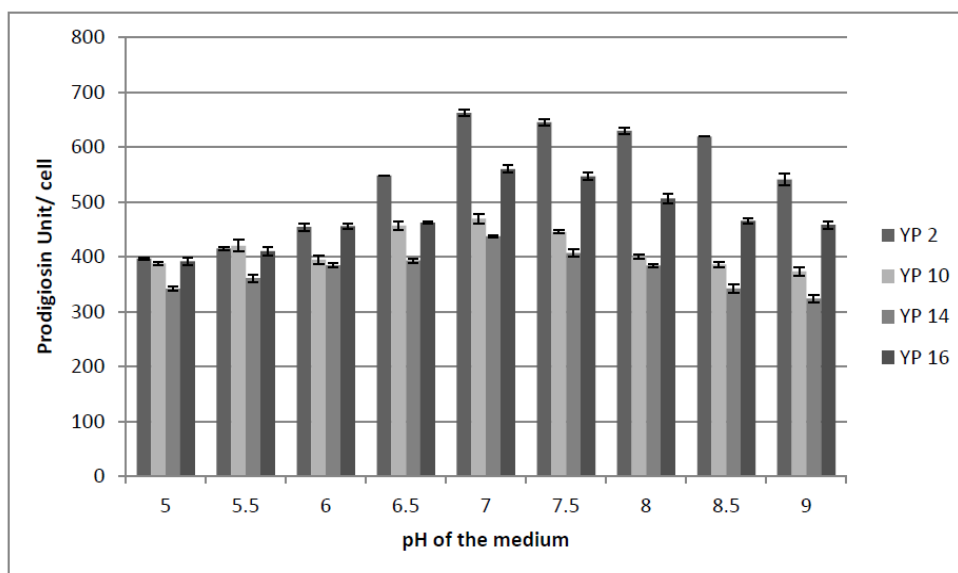
**RESULT AND DISCUSSION****Optimization of production process.****Effect of pH of the medium on prodigiosin production.**

The pH plays a vital role in maintaining the acidic or basic condition in the medium. The pH provides an environment for the microorganism to grow luxuriantly.

Effect of different pH viz. 5, 5.5, 6, 6.5, 7, 7.5, 8, 8.5, and 9 was tested on production of prodigiosin. After incubation pigment was extracted and the concentration was estimated. The maximum yield of pigment in unit/cell (662.1), was obtained in the peanut broth with pH 7 when inoculated with YP 2. Followed by YP 16, YP 10 and YP 14. Further increase in pH resulted in decreased in prodigiosin production. Pryce, L. H. *et al.*, (2000) reported that, more prodigiosin was produced at pH 7. Davaraj, N. *et al.*, (2009) have used trypticase soya broth for the production of prodigiosin. They have tested pH 4 to 10 for the optimized prodigiosin production. They found that pH 7 and pH 8 shows maximum prodigiosin production. Sundaramoorthy, *et al.*, (2009) was found that, prodigiosin production was maximum at pH 7.0. Our result also in agreement with result obtained by Chandni, *et al.*,(2012). Previous studies also reveal that the glucose decreased the yield of pigment when present in the medium due to lowering the pH (Solo, *et al.*, 1997).

**Table No.1: Effect of pH of the medium on Prodigiosin production.**

pH of Medium	YP2	YP10	YP14	YP16
5	396.89 ± 1.95	387.81 ± 3.44	342.01 ± 4.34	391.70 ± 7.06
5.5	415.3 ± 2.73	419.97 ± 10.83	361.11 ± 7.08	410.12 ± 7.68
6	454.55 ± 6.76	394.82 ± 8	384.67 ± 3.67	455.87 ± 5.43
6.5	548.45 ± 0.23	456.94 ± 8.02	393.50 ± 3.94	462.47 ± 2.16
7	662.1 ± 5.68	469.58 ± 9.22	437.67 ± 1.67	560.25 ± 6.02
7.5	645.34 ± 5.21	444.99 ± 2.94	406.96 ± 6.14	546.86 ± 6.57
8	629.42 ± 5.01	400.87 ± 4.50	384.10 ± 3.28	506.22 ± 8.45
8.5	619.43 ± 0.27	386.03 ± 5.22	342.64 ± 7.88	465.95 ± 5.14
9	541.02 ± 10.51	373.49 ± 7.98	323.76 ± 7.63	458.58 ± 6.79

**Fig. No. 1: Effect of pH of the medium on Prodigiosin Production.**

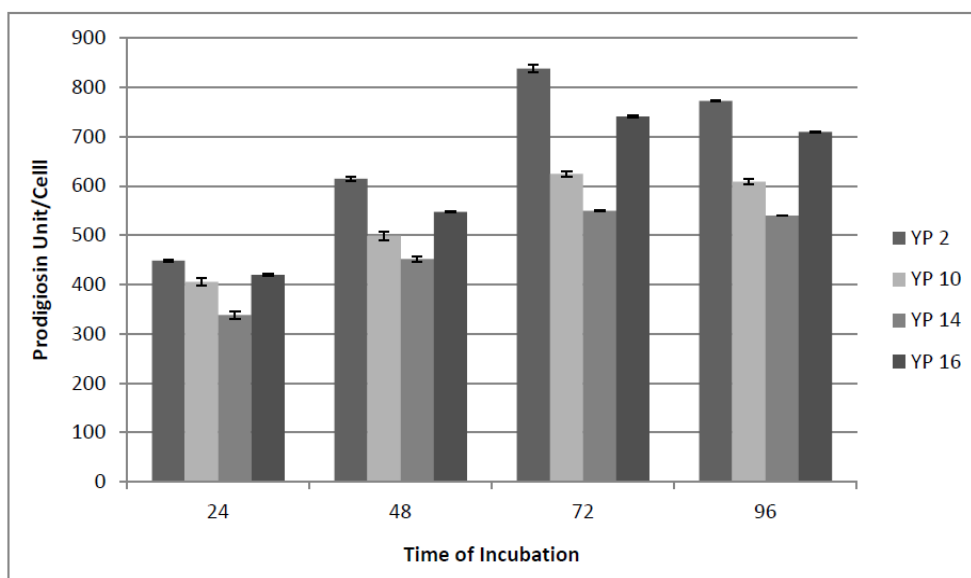
### Effect of time of incubation on prodigiosin production.

Effect of different incubation time viz. 24, 48, 72 and 96 hr. on production of prodigiosin were tested. After equal time interval pigment was extracted and the concentration was estimated. The maximum yield (838.28) of pigment in unit/cell was obtained after 72 hr. incubation when inoculated with YP 2. Followed by YP 16, YP 10 and YP 14. Further increase in incubation time do not showed notable effect on pigment yield.

Davaraj, N. *et al.*, (2009) has reported maximum pigment production on fifth and seventh day of incubation whereas the pigment production completely reduces after seventh day of incubation. Samrot, *et al.*, (2011) reported that, more prodigiosin was produced after incubation of 72hr. After 72 hr. the rate of prodigiosin production was reduced. The observation suggests that producer organism produced pigment in stationary phase of growth.

**Table No. 2: Effect of time of incubation on Prodigiosin production.**

Time (hrs)	YP2	YP10	YP14	YP16
24	448.72 ± 2.13	405.55 ± 7.73	338.15 ± 8.20	419.76 ± 2.46
48	614.3 ± 4.94	498.72 ± 8.80	452.05 ± 5.61	547.14 ± 1.06
72	838.28 ± 7.84	624.39 ± 5.38	549.68 ± 1.05	740.81 ± 2.62
96	772.76 ± 1.07	608.32 ± 5.34	539.78 ± 0.53	709.3 ± 0.34



**Fig. No. 2: Effect of Time of Incubation on Prodigiosin Production.**

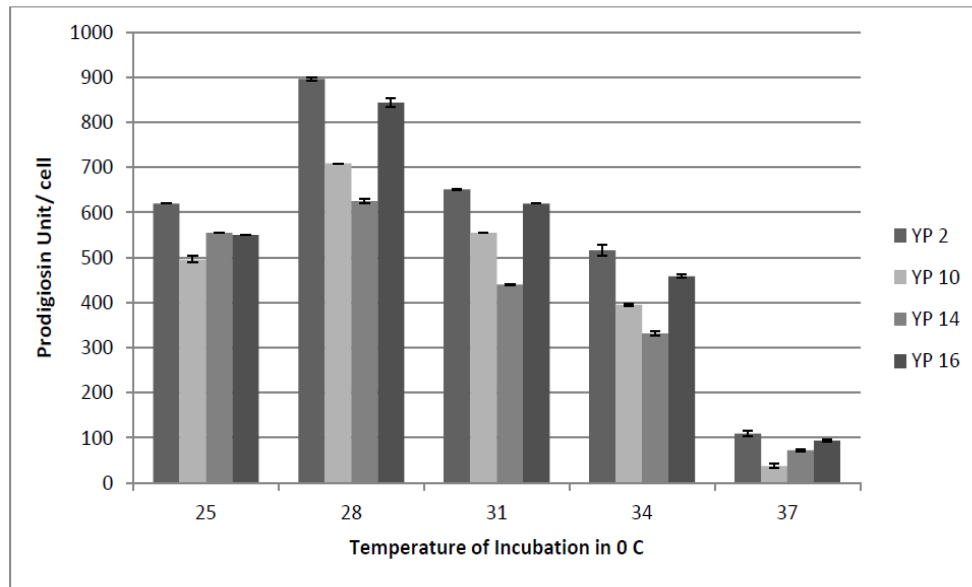
### Effect of temperature of incubation on prodigiosin production.

Effect of incubation temperature viz. 25, 28, 31, 34 and 37°C on production of prodigiosin was studied. After incubation, pigment was extracted and the concentration was estimated. The maximum yield (895.90) of pigment in unit/cell was obtained in 28°C when inoculated with YP 2. Followed by YP 16, YP 10 and YP 14. It was also found that with increase in incubation temperature, there was a decrease in the yield of prodigiosin.

According to Samrot, *et al.*, (2011) prodigiosin not produced above 37°C, but as temperature decreases rate of production increases. The organism was found to produce more prodigiosin at 28°C at 72 hr. Williams & Hussain Quadri (1980) reported that, no prodigiosin was produced when cultures were incubated at 38°C, however pigment production was observed when the temperature was shifted to 25°C -30°C.

**Table No. 3: Effect of incubation temperatures on Prodigiosin production.**

Temperatures °C	YP2	YP10	YP14	YP16
25	619.34 ± 0.32	495.59 ± 6.94	554.5 ± 0.60	549.86 ± 0.92
28	895.90 ± 3.38	708.41 ± 0.53	625.10 ± 5.42	843.91 ± 9.39
31	651.21 ± 0.31	554.46 ± 0.60	439.62 ± 1.39	619.01 ± 00
34	515.63 ± 11.86	394.91 ± 2.96	331.45 ± 4.08	458.49 ± 3.00
37	108.96 ± 5.52	36.87 ± 5.06	71.58 ± 3.14	93.14 ± 2.11



**Fig. No. 3: Effect of incubation temperatures on Prodigiosin production.**

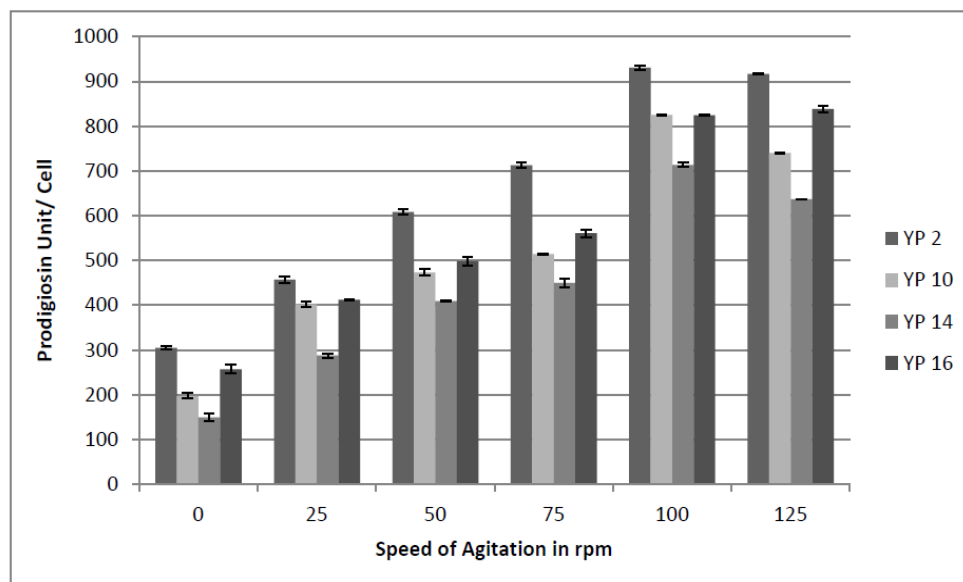
#### Effect of agitation on prodigiosin production.

Effect of speed of agitation (viz. 0, 25, 50, 75, 100 and 125 rpm) on production of prodigiosin was determined by incubating inoculated peanut broth at various rpm. After production of prodigiosin it was extracted and the

concentration was estimated. The maximum yield (930.74) of pigment was obtained in flask agitated at 100 rpm when inoculated with YP 2. With the increase in the speed of the agitation above 100 rpm, a decrease in the yield of prodigiosin was observed.

**Table No. 4: Effect of speed agitation on Prodigiosin production.**

rpm	YP2	YP10	YP14	YP16
0	305.12 ± 4.48	197.22 ± 5.80	149.73 ± 7.64	256.85 ± 10.11
25	456.94 ± 8.02	401.88 ± 6.25	287.42 ± 5.31	412.14 ± 1.61
50	608.22 ± 5.88	473.26 ± 6.60	408.81 ± 1.70	498.24 ± 8.77
75	713.29 ± 5.16	514.27 ± 0.75	449.5 ± 8.89	560.8 ± 8.42
100	930.74 ± 4.49	825.4 ± 1.41	713.62 ± 4.97	824.39 ± 0.60
125	916.35 ± 1.55	740.10 ± 0.80	636.44 ± 0.55	838.28 ± 7.84



**Fig. No. 4: Effect of Speed of agitation on Prodigiosin Production.**

## Effect of light spectrum on prodigiosin production.



Fig. No. 5: Experimental setup to check effect of light spectrum on production process.

Table No. 5: Effect of Light Spectrum on Prodigiosin Production.

Time (hrs.)	Black	Blue	Green	Yellow	Red	White
24	331 ± 40.0	257 ± 5.57	158 ± 11.7	240 ± 41.5	200 ± 43.8	392 ± 4.7
48	470 ± 8.7	549 ± 30.4	279 ± 32.3	427 ± 35.25	338 ± 5.8	534 ± 23.1
72	602 ± 9.4	650 ± 9.2	607 ± 5.6	624 ± 4.9	521 ± 27.7	816 ± 9.8
96	560 ± 5.8	624 ± 5.5	549 ± 6.1	581 ± 19.5	478 ± 5.4	599 ± 10
120	514 ± 13	360 ± 7.3	284 ± 21.6	540 ± 40.3	258 ± 4.5	556 ± 5.5

## Effect of light spectrum on pigment production (unit / cell).

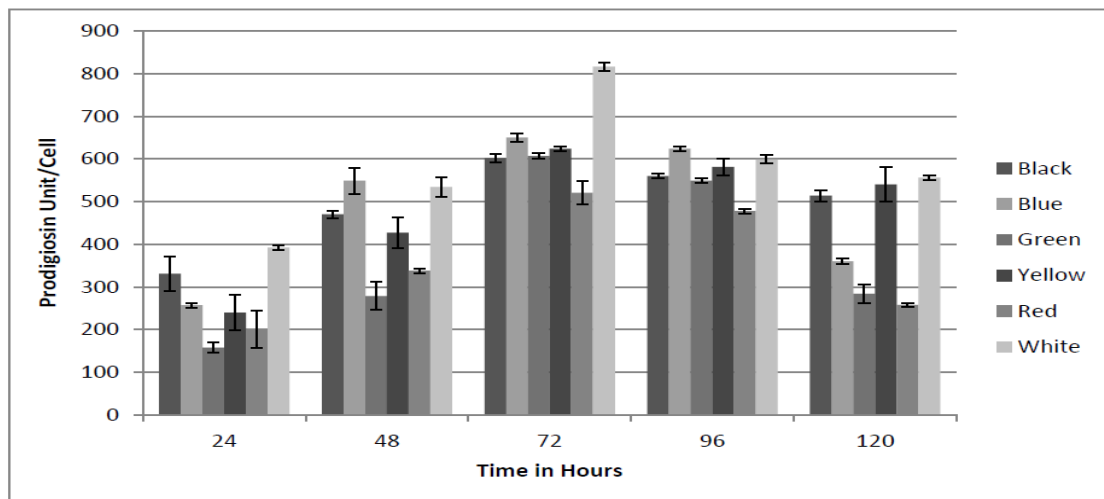
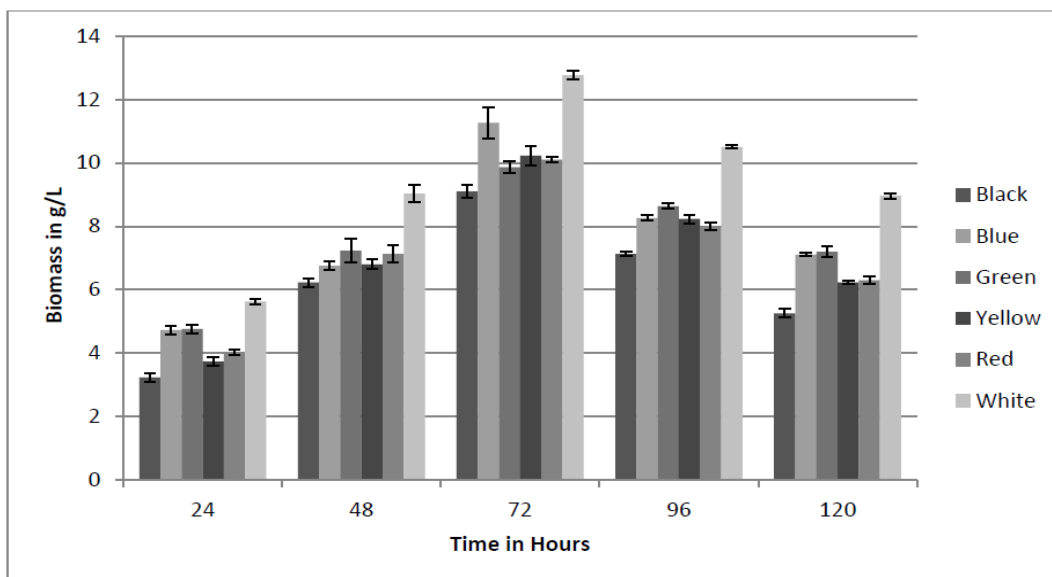


Fig. No. 6: Effect of Light Spectrum on Pigment Production.

## Effect of light spectrum on biomass production (g / L).

Table No. 6: Effect of Light Spectrum on Biomass Production.

Time (hrs.)	Black	Blue	Green	Yellow	Red	White
24	3.23 ± 0.14	4.73 ± 0.14	4.76 ± 0.14	3.73 ± 0.14	4.03 ± 0.08	5.63 ± 0.08
48	6.23 ± 0.14	6.76 ± 0.14	7.23 ± 0.37	6.8 ± 0.15	7.13 ± 0.27	9.03 ± 0.26
72	9.1 ± 0.20	11.26 ± 0.50	9.86 ± 0.18	10.23 ± 0.31	10.1 ± 0.09	12.76 ± 0.14
96	7.13 ± 0.08	8.26 ± 0.09	8.63 ± 0.08	8.23 ± 0.14	8 ± 0.11	10.5 ± 0.05
120	5.26 ± 0.14	7.1 ± 0.05	7.2 ± 0.18	6.23 ± 0.05	6.3 ± 0.11	8.96 ± 0.08



**Fig. No. 7: Effect of Light Spectrum on Biomass Production.**

The results show that production of pigment and biomass in *Serratia spp.* is highly depends on the intensity and wavelengths of light. Here, the culture under submerged condition was exposed to different colors and thus different wavelengths of light (Blue, Green, Yellow, Red, White and Darkness). Maximum pigment production was observed in white light  $816 \pm 9.8$  unit/cell followed by blue light ( $650 \pm 9.2$  unit/cell) and yellow light ( $624 \pm 4.9$  unit/cell) (Fig. No.6), though incubation of organism in black, red and green light gives comparatively less yield, it was not completely inhibited. Increased pigment production was observed in total white light as compared to the flasks completely exposed to darkness. The visual comparison of the extracted pigment exhibited varied shades of color to different wavelengths of light. The white light incubation yielded a deep red colored pigment whereas yellow light

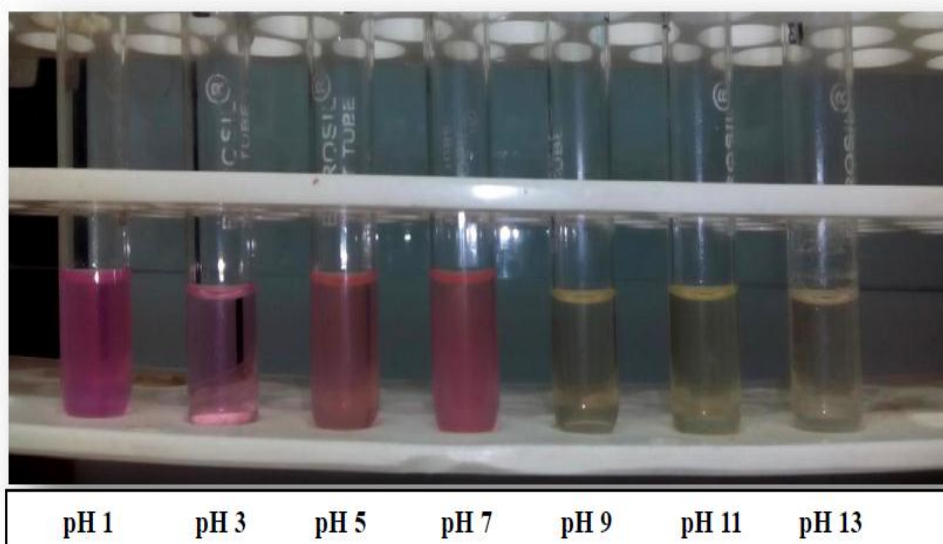
incubation yielded a light red to orange colored pigment. In relation with the above observation white, blue and yellow light give high yield of biomass ( $12.76 \pm 0.14$ ,  $11.26 \pm 0.50$  and  $10.23 \pm 0.31$  respectively), whereas in red and green light biomass yield was decreases (Fig.7). The growth was also monitored on solid media to determined effect on colony diameter and morphology. Colony exposed to White light exhibited highest radial growth and red light showed the least diameter on the solid media. No notable effect of other lights on the said character was observed.

The effect of light source on the pH of the medium was also determined and it was found that as time of incubation increases pH of the medium also increase from neutral to basic in all flasks.

#### Stability of pigments to different pH.

**Table No.7: Stability of Prodigiosin to pH.**

Sr. No	pH	Colour	Lambda Max (nm)
1	1	Dark Pink	580
2	3	Pink	560
3	5	Light Pink	550
4	7	Reddish	540
5	9	Yellow	530
6	11	Light Yellow	460
7	13	Colorless	400



**Fig. No. 8: Stability of Prodigiosin to pH.**

At extremely acidic pH (1 to 5), the red colour pigment appeared as pink probably due to protonation of pyrrole ring nitrogen facilitated by high concentration of H<sup>+</sup> ions in the solution. While in highly basic pH (9 to 13), the pigment was almost decolorized. The decrease in color intensity in basic condition can be attributed to the deprotonation of nitrogen from the three conjugated rings of pigment structure that give it stability (Komzen, *et al.*, 2006). The UV visible analysis of each tube gives maximum absorption peak ( $\lambda_{max}$ ). For this methanol was used as blank.

It was found that absorption maxima changes with pH. The primary reason for stronger absorption of pigment at lower wavelength was the electron conjugation effect or delocalization of electrons, which is substantiated by Mohan, *et al.*, 2007.

## CONCLUSION

In the present study it was concluded that peanut seed medium was a good medium for prodigiosin production. Out of selected four strain of *Serratia spp.* YP 2 was found as potent pigment producer when inoculated in the medium with pH 7, time of incubation 72hr. and temperature of incubation 28°C. It was also found that agitation of the medium can significantly affect production process. The study also reveal the effect of different wavelength of light on pigment and biomass producing ability of *Serratia spp.* It shows that white light is best for pigment and biomass production followed by blue and yellow light. The red and green light showing inhibitory effect on the pigment production. As pH of the medium increases linearly with the time, indicating that, at the time of growth and production process of the pigment, producer organism does not produce acids in the medium. The stability of produced pigment to the pH was also check, which shows that pigments coloration pattern is very much sensitive to change in the pH of the medium.

From the result obtained it was concluded that production of pigment and biomass in *Serratia spp.* is greatly depend upon type of light. The effect of intensity of light spectrum on the red pigment production can be a topic of interest in future for the researcher working in this area. The color of the pigment and its lambda max value changes with change in pH, suggest that pigment prodigiosin is very much sensitive to pH change and it can be used as pH indicator.

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