

CRISPR CAS9: A NEW TECHNOLOGY TO MODIFY GENOME- A REVIEWPankaj Bhatt*¹, Suruchi Singh², Narjes Alfuraiji³, Ali E. Al-Snaffi⁴¹Department of Pharmaceutics, KIET Groups of Institutions (KIET School of Pharmacy), Muradnagar, Ghaziabad, U.P, India.²Department of Pharmacology, Glocal University, Saharanpur, U.P, India.³Department of Pharmacology, College of Medicine, University of Karbala, Iraq.⁴Department of Pharmacology, College of Medicine, University of Thiqr- Iraq.***Corresponding Author: Pankaj Bhatt**

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ABSTRACT

The simplest, cheapest and most valuable method of modifying the genome was CRISPR technology, which assists the scientist in gathering the DNA chain and reforming its volume. CRISPR is a most significant prospective implementation against genetic imperfection to develop and protect assortment from broadening of illness. Different catalogue databases were previously published arrange, audit papers traverse, and systemic data completed in a different CRISPR implementation. Look into various implements and effects of gene editing types of equipment and other areas. Likewise, it helps to improve benefits and virtue of matter related to CRISPR Cas9 abbreviation was CRISPR. The technology has been modifying the resistance mechanisms or unicellular organisms such as bacteria. The organisms employ the CRISPR inferred RNA protein, which deducts the virus's capability to attack. Di action can do to destroy DNA from invading by far apart. It permits the constituents to relocate monitor and the changing of the genes. An extensive review of seventy-six papers involving research and review works was done to let the work become quickly, and straightforward forward valuable and unique restriction Method of standard modern and recently developed system to cure illness and scrutinise. Even though various current therapeutic systems and different therapeutic system, were available, gene editing technology was available to cure cancer, Ocular, AIDS, muscular dystrophy, tons illness, blood disorders, and others that reduce pain. Furthermore, they intermingled with some particular side effects. To resolve the problem of drug delivery, and it shows efficacy by overcoming the side effects; however, nevertheless, the famous and safety well-being have been the significant hindrances against their Victorious location.

KEYWORDS: CRISPR, Cas9, Gene editing, Genetic defects, Genome engineering, Gene driving.**1. INTRODUCTION**

CRISPR is the type of technology for gene editing that allows scientists to cut and paste and turn genes into DNA. It is concerned with the DNA destroying defence system that is found in some prokaryotic. CRISPR dominates in gene editing. The first gene editing performed by CRISPR was calculated as 6.8. There are many exploded cases that are being done by CRISPR since 2013. Short form for CRISPR is clustered regularly interspaced short palindromic repeats. The nuclease associated with CRISPRs is Cas9.^[1-3] The 29-nucleotide repeat separated with a 32-nt spacer sequence was first shown in bacteria in 1987. Subsequently, 40% was found in the sequence bacterial genome and 90% in archaea.^[4] Many types of cash jeans are well deserved and are adjusted to repeated elements.^[5] In 2005, the spacer sequence was found in phage genomes, which was considered the year of memorable CRISPR.^[6] The full form of CRISPR describes the group of nucleic acid

sequences discovered in bacteria and was discovered by Archaein 1990 that contains several copies of virus genes.^[7-9] The various copy-edit methods were zinc finger nuclease transcription activator-like effect or nucleases and the CRISPR-related system Cas9 every time the DNA sequence more so. It comprises only one guide RNA molecule whereby the earliest 20 of the nucleotide harmonises to the DNA target,^[10,11] so Cas9 every time produces DSBs at a specific position by the time RNA existence in genomic DNA.^[12]

BSB originated from CAS9 and for the particular RNA, cellular DNA is found to mix with DSB for machine repair.^[13] If the error is found in the non-homologous end joining DNA system used for this process, mistakes are also made and rectified and introduced, leading to disruption of gene function.^[14-18] The snip DNA fragment is stored between the palindromic CRISPR sequence to preserve the available memory to prevent

future infection from the previous viral strain.^[18] In this, we will explain how CRISPR opens a new structure for cancer research.

What the heck is CRISPR?

To understand CRISPR in a much better way, we need to study about 1987, when Japanese scientists used to study *E. coli* bacteria, the first sequence found and repeated in the organism of DNA was discovered and repeated.^[20,21] The biological importance and role of the sequence are not known.^[22] As time passed, many more scientists have found about it in the same manner for other bacteria's DNA.^[23] A proper name is given to this sequence that is Clustered.^[24] Regularly Interspaces Short Palindromic Repeats. However, the role of CRISPR is still not yet known till 2007.^[25] By studying *Streptococcus* bacteria, it is used in yoghurt and plays a vital role.^[26] Bacteria are at constant hit from viruses and produce enzymes that help fight it.^[27] Different viruses arise when killing the virus; it scoops the leftover virus code and breaks it into tiny pieces. After which, it is store in the space of CRISPR in the bacterium genome. CRISPR also works like a devil for viruses and bacteria that use genetically saved cells to defend themselves against the next attack.^[28] A particular attack enzyme is produce called CRISPR associated protein 9 (Cas9) at new viral infection.^[29]

How CRISPR is used

CRISPR-based technology has been used for tasks in recent years, whether related to the killing drug-resistant superbugs or removing the responsibility of gene for the disease or creating a molecular recording device. Concerns about risks and ethics and ethical issues related to their application were raised. At last of the year 2018, Chinese scientist He Jiankui said that he has been using CRISPR that is being carried in both and are still given for investigation.^[30]

CRISPR application

- CRISPR in gene therapy and medicine
- Diseases CRISPR technology can cure

Scientists are dealing with eight diseases. With the help of CRISPR Cas9, it has become the primary condition required to treat revolutionary technology.

Cancer

The common primary disease associated with increased mortality that is rising throughout the world is cancer. CRISPR application is made first on cancer.^[31] The first advanced CRISPR clinical trial is being conducted in China that can test the ability of gene editing tools that are useful for treating patients with the esophagus.^[32] The testing is being done at Hangzhou Cancer Hospital, beginning with the extraction being done with the patient's immune T cells. Using CRISPR says it is arranged to replace the gene encoded with the protein called PD-one few tumours can bring together the protein on the surface of immune cells and stop them

from attacking.^[33] They replace LR after being re-infused in the patient with a higher ability to attack cancer cells.^[34]

CRISPR - A versatile tool for genome engineering

CRISPR and CRISPR-associated proteins are grouped up to target the foreign viral DNA to the adaptive immunity.^[35] The two different Macanese RNAs - CRISPR targeting and trance activating RND - activate and help guide the CAS protein for binding viral DNA sequences claimed.^[36-38] Likewise, proteins induce some single nucleotide base changes that Cas9 variants.^[39]

Design of SgRNAs and bio information resources

Like earlier techniques and technology, CRISPR is being used as a genome editing tool spread throughout the scientific community.^[40] It is being supported by Developing and designing through open bioinformatics resources and analysing that is done through CRISPR associated experiments. Recently, there are different types of online tools available for RNAs related to the efficiency and quality being measured.^[41] The system that helps to find RNA focusing on its efficiency is calculated from extensive screening data and integration through nucleotide composition and its position within the gene model.^[42,43]

Target discovery by CRISPR/CAS9 screens

An essential tool used for cancer therapy treatment discovers that a novel target is CRISPR.^[44] For CRISPR to be generated, a large variety of Geneknock outs is required. The third requires a variety of practical steps and bioinformatics. For every target gene to be started, the primary step is to predict the efficiency of sgRNAs.^[45,46] Different CRISPR/Cas9 screens technologies are used to identify between two functional entities. It was found that in the genome CRISPR separated pancreatic cell lines in E3 Ubiquitin ligase ring finger protein 43, especially endangered to eliminate the Wnt ligand (a comprehensive family of release glycoproteins) receptor frizzle-5(FZD5). The standard that is being merged has been forecast since, in the algorithm, the small synthetic interlink of dissimilar CRISPR separation was conducted.^[47]

Interrogation of the non-coding genome of cancer

CRISPR has proved to be a remarkable tool that is used to interrogate non-coding elements. Three cancer-associated genes, Cullin 3, Neurofibromatosis type 1, and Neurofibromatosis type 2, help identify the enhancer region.^[48] It is being performed for the genomic region of about 700 KB that surrounds the candidates' complete reading frame. CRISPR requires one more piece of information to identify target genes.^[49] For this reason, the genomic region, despite sgRNA, shows both TP53 binding property and enhancer Markers. It was hitting off a required element result for the resistance of cells against HRASG12V induced senescence. It is connected to TP53. Multiple enhancers in the presence of Cyclin-dependent kinase inhibitor 1A (CDKN1A), which are

found on editing, have been identified to result in the down-regulation of Cyclin-dependent kinase inhibitor 1A (CDKN1A) and due to which it can easily escape from oncogene-induced senescence. Non-coding RNAs are another class of poorly genetic elements. In general, three different CAS9 techniques were systematically used to detect the role of long code RNA in cancer cells. Zhu *et al.* used pairs of sgRNAs to delete 651 long noncoding RNA genomic sequences. Subsequently, 51 non-coding long RNAs were changing the Proliferation phenotype after elimination.^[50]

Blood disorders

The primary CRISPR trial was started in Europe and the US to treat the patient in February for beta-thalassemia and sickle cell infection. The two blood problems influence oxygen transport in the blood. It involves the immature microorganisms of the bone marrow of the patient and the CRISPR innovation created by CRISPR Therapeutics and vertex drug. It generates a vital oxygen-carrying protein, which brings oxygen together better than in the adult form. Another type of blood disorder in which CRISPR technology can work is hemophilia. The technology works with casebia employing CRISPR in which the tool for gene editing is being conveyed directly to the liver.

Eye-related genetic diseases

Treating genetic blindness CRISPR is known as the best technology. The hereditary form of blindness, which is due to a particular mutation, makes it easier for CRISPR-Cas9 to target and modify a single gene.^[51] The eye is an immune-privileged part of the body, so it is minimal here, which is an advantage of Insight that shows the possibility that CRISPR ensures the reaction against it that can drive and produce side effects. The main reason for childhood birth blindness is Lebercon genital amaurosis. The medicine that is continuously progressing for CRISPR therapy is Editasmedicine. For this, there was no treatment.^[52] The company's primary purpose is to ensure the light-sensitive cell's functioning so that children do not lose their vision. The most important therapeutic target used in the case of diseases is VEGFR2.^[53]

Liver-related genetic diseases

The treatment option used to cure liver-related diseases like hereditary tyrosinemia is the correction of genes for the CRISPR system.^[54] System delivery of CAS9 mRNA dealing with the C 12–200 lipid in peace and the HDR template dealing with an AAV vector help correct approximately 6% of hepatocytes found in the mouse model of human hereditary tyrosinemia.^[55,56] Groups are being developed to deliver CRISPR components to the liver. BAMEA-016B And TT3 are the two MPs used for systematic delivery of CAS9-mRNA, and gRNA Are the mice liver that allows the PCSK9 gene to edit and for the regulation of PCSK9 protein level.^[56]

AIDS

CRISPR technology plays an important role in fighting against AIDS. One way is to remove the DNA from the hiding place to the DNA in the immune cells. This method is useful for attacking the virus, which did not place an inactive form, making it suitable for maximum therapies to fight against viruses. Another way through which the virus can be removed is for resistance to HIV infection. Few people from birth have resistance of HIV to a mutation in a gene that is particularly called CCR5, which ensures that HIV is used as the entry point for infecting cells. Not allowing the virus to hold it together mutation helps to change the complete structure of the protein. Last year in China, this method is found in a controversial case. For editing the embryos to make them resistant to HIV infection, CRISPR-Cas9 plays an important role. The researchers and experimenters thought among the community that the baby with CRISPR has a greater risk that they may die earlier. The consensus seems that more and more information and study are being required before using it for humans.^[57]

Cystic fibrosis

Cystic fibrosis is responsible for some respiratory problems. The person with this disorder has an age limitation of only 40 years after being treated. By editing the mutation that causes cystic fibrosis, CRISPR technology plays a vital role in finding the necessary foundation in the CFTR gene. Scientists have shown that CRISPR can be induced in humans, found in patients with cystic fibrosis that fixes the disease mutation. The next procedure is to test it in humans, both in medicine and in CRISPR therapeutics.^[58] There are many different ways of mutation In the CFTR Jean for cystic fibrosis. This means that different areas of CRISPR therapy are used to develop different genetic defects.^[58]

Muscular dystrophy

In general, the money is that Duchenne muscular dystrophy is responsible for mutations in the DMD diastrophic gene responsible for the suitable protein for muscle contraction. Mainly the patient who suffers from this disease, there is a lot of muscle collapse, and there is no possible cure for this disease, yet research done in mice proves that CRISPR technology is used to correct many genetic mutations found in the disease. Many researchers in the USA have found the new idea instead of the previous one used by CRISPR technology to remove or reduce 12 strategic mutation hotspot that covers a maximum of 3000 different mutations responsible for the muscular disease. A company named exonic therapeutics is responsible for developing this method.^[59]

Huntington's disease

The neurodegenerative condition that has a substantial and powerful genetic component is Huntington's disease. When there is an abnormal repetition of DNA sequences found in the Huntington gene, this disease is caused. Treatment of this disease is complicated as it can be

dangerous for the brain. Scientists are looking for methods and techniques to edit the gene tool to make it easier and safer.^[60]

CRISPR in other fields

CRISPR – A new tool for gene driving technology

CRISPR clustered regularly into space-short Palindromic repeats, an editing tool that was being developed in 2011. In this tool, Arena helps bring the Indo nucleus to a specific target with ease. The Endonuclease Cleaves that DNA targeted sites are triggering the cell for repairing the double-strand breaks. The repair process makes changes found in the target site.

For repairs, the cell mainly uses variants present in the chromosome in a process called HDR. After repairing, the drive allele is copied to the wild type of chromosome, removing wild type DNA sequences with the genome.^[61]

CRISPR has expanded transgenic animal research

The CRISPR-CAS9 technology that is being developed in 2012 has changed entirely in genetic engineering. As CRISPR enables the editing of targeted genomes in a simple, efficient, and economical manner, the process of creating transgenic animals became simpler in the span of just a few years. Moreover, the commercial availability of synthetic single-guide RNA reduced the experimental time and improved the editing efficiency compared to in vitro transcribed guide RNA. For instance, Synthego Synthetic RNA shows high editing efficiencies and a high germline transmission rate, which are critical factors for successfully generating transgenic animals. The development of CRISPR based on nonengineering technology edits the gene in such a more straightforward way that creates transgenic mice in the lab. They are now available to guide and instruct the researchers and scientists.^[62]

Application of CRISPR/CAS9 in plant genome editing

The application of CRISPR in the plants is developing. In Arabidopsis, a new plant with various genes consisting of ATFLS2, ATSP4, ATPDS3, etc., they are targeted with the different mutational efficiencies ranging from 1.1% to 84.8% in the first generation.^[63-69] These are heritably found in a variety of generations and a percentage rate of 79.4.^[69] For modifying the two genes, a single CRISPR with two different expressional cassettes is being developed. In tobacco, CRISPR is being joined together with version VIGS technology, and the internet to it causes morphological changes found in the compound leaves.^[70-72] CRISPR helps in editing, and its use has been developed from time to time.^[73]

CRISPR applications in crop genetic improvement

The CRISPR system is straightforward, adequate, efficient, simple, and highly specific, helping to produce target events. However, it is an essential tool that plays a crucial role in the modification of the genome in plants. For basic and higher levels of plant biology research,

CRISPR has shown an important role. It also has an impact on crop breeding. The editing of genomes helps get the modification in a session that saves time and is simple for the conventional breeding scheme. CRISPR also proves the best approach for pyramid breeding.^[74]

CRISPR in de-extinction

Even though an idea like this was brought from creative science writing, it was not scientist that was running on. Since to get back animal which wire varnished the initial prospects of pigeon passenger the moment of notes American green wood resident employee. CRISPR techniques investigators schedule to initiate gene through the gene of pigeon passenger to recent present time comparable the stripe back and of pigeon, the hybrids will have been bread for many creations till DNA progeny resembles that the varnished genus of the initial creation of resuscitative pigeon was anticipated to generate in the year of 2020.

More nutritious fish

The genesis customises Salman fish which raise two fold rapidly were standard are yield in Canada. When the CRISPR arrived, gene-editing scientist would have promoted an investigation group, presently CRISPR-CAS9 was employed to produce purified Salman, which allowed him to increase in number better and become mota less likely to suffer from illness. Furthermore, it halts the Salman reproduce sexually it control flee the Aquaculture provision the investigators currently research the characteristic that would strengthen CRISPR use. We were so excited to make the genesis more comforts for the fish as well as resistance to disease. Anna Marsilius head of analysis group, spoke to me that the technology could be used also to emphasise the omega-3 parliament of the fish in order to make well-being and even more famous in the upcoming days.

Eradicating pests

We used to maintain the spread of infectious disease; even an animal species figure capable of doing so, affecting the biological community of interacting organisms and their physical environment by using CRISPR. The gene-editing technology employs to generate gene-drive which providing the genetic alteration would take over by all progeny open out all over the habitat numbers throughout the many creations. A year later, scientist forum UK Imperial College London confirm that the technology can also help the mosquito genus, which is in charge of transmission of paludism (malaria). Which initiates gene once both parents of them take it the process of lay in the egg by female anopheles mosquitoes would have been halted in the experiment with the lock-up mosquito was cleaned up once number create later on gene drive has been initiated.^[75]

Clinical applications of CRISPR

Long before developing a gene-editing technique, this has been talked about editing the cell culture model's

genes, organism, and humans.^[76] Traditionally in this gene therapy, the new genetic material edited is inserted into somatic cells, after which the edited gene is shown to be effective in curing disease by displaying its therapeutic effect.^[77] Gene therapy trials have been in place since the 1980s, but most experiments have been conducted over the years due to somatic cells not reacting appropriately with genes, the mutual immune response to the host, and changing genetic information have failed. Clinical trials first published in 2014 confirmed the possibility of gene editing technique in the right direction but did not use the zinc finger technique. In this test, the T cell present in mice was targeted by ZFN using the HIV co-receptor. This concept, as confirmed by a study done by the chain *et al.* We have been handed over the general judgment from this aimless review article to target mutagenesis and numerous gene-editing CRISPR employ in different sectors. Editing role or swap gene pronouncements The CRISPR-Cas9 genome editing technique can be employed via the different method through mixing it with one more gene editing technique the CRISPR/Cas9 technique is fundamental and translational Cancer research was valuable since which has extreme utility in the upcoming time. In the coming years, pooled CRISPR screening technology will bring forth an additional extensive set of the critical gene, which will play a vital function again cancer cells. The time of genetic engineer in demand was increasing rapidly for prophylactic of disease, and therefore we should keep an eye on the extensive influence for this field to remain in genetics as a starting point of understanding the therapeutic area of the gene.-

CONCLUSION

This review article presented the general evolution from random to targeted mutagenesis and various genes editing (CRISPR) used in different fields. CRISPR-Cas9 is based on CRISPR-associated proteins and these can perform genome editing functions or change gene expression. The CRISPR-Cas9 genome editing technique can be used via a different method by combining it with another gene editing technique. The CRISPR / Cas9 technique in fundamental and translational cancer research was valuable, which has extreme utility in the upcoming time. In the coming years, pooled CRISPR screening technology would bring forth an additional extensive set of the vital gene, which will play a vital function again cancer cell lines. By the time of genetic engineering, demand was increasing rapidly for prophylactic of disease; by then, we should keep an eye on extensive influences for this field to remain in genetic as starting point of human disease and comprehend the therapeutic area of gene.

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Conflict of interest

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REFERENCES

- Cong L, Ran FA, Cox D, Lin S, Barretto R, Habib N, *et al.* Multiplex genome engineering using CRISPR/Cas systems. *Science*, 2013; 339(6121): 819–823.
- Mali P, Yang L, Esvelt KM, Aach J, Guell M, DiCarlo JE, *et al.* RNA-guided human genome engineering via Cas9. *Science*, 2013; 339(6121): 823–826.
- Jinek M, Chylinski K, Fonfara I, Hauer M, Doudna JA, Charpentier E. A programmable dual-RNA-guided DNA endonuclease in adaptive bacterial immunity. *Science*, 2012; 337(6096): 816–821.
- Ishino Y, Shinagawa H, Makino K, Amemura M, Nakata A. Nucleotide sequence of the IAP gene, responsible for alkaline phosphatase isozyme conversion *Escherichia coli*, and identification of the gene product. *J Bacteriol.* 1987; 169(12): 5429–5433.
- Horvath P, Barrangou R. CRISPR/Cas, the immune system of bacteria and archaea. *Science*, 2010; 327(5962): 167–170.
- Jansen R, van Embden JDA, Gaastra W, Schouls LM. Identification of genes that are associated with DNA repeats in prokaryotes. *Mol Microbiol*, 2002; 43(6): 1565–1575.
- Bolotin A, Quinquis B, Sorokin A, Ehrlich SD. Clustered regularly interspaced short palindrome repeats (CRISPRs) have spacers of extrachromosomal origin. *Microbiology*, 2005; 151(Pt 8): 2551–2561.
- Mojica FJM, Díez-Villaseñor C, García-Martínez J, Soria E. Intervening sequences of regularly spaced prokaryotic repeats derive from foreign genetic elements. *J Mol Evol*, 2005; 60(2): 174–182.
- Pourcel C, Salvignol G, Vergnaud G. CRISPR elements in *Yersinia pestis* acquire new repeats by preferential uptake of bacteriophage DNA and provide additional evolutionary studies tools. *Microbiology*, 2005; 151(Pt 3): 653–663.
- Mahfouz MM, Li L, Shamimuzzaman M, Wibowo A, Fang X, Zhu J-K. De novo-engineered transcription activator-like effector (TALE) hybrid nuclease with novel DNA binding specificity creates double-strand breaks. *Proc Natl Acad Sci USA*. 2011; 108(6): 2623–2628.
- Mishra R, Joshi RK, Zhao K. Genome editing in rice: Recent advances, challenges, and future implications. *Front Plant Sci.*, 2018; 9: 1361.
- Wu X, Kriz AJ, Sharp, PA. Target specificity of the CRISPR-Cas9 system. *Quant Biol.*, 2014; 2(2): 59–70.
- Liu L, Fan X-D. CRISPR-Cas system: a powerful tool for genome engineering. *Plant Mol, Biol.*, 2014; 85(3): 209–218.

14. Ma X, Zhu Q, Chen Y, Liu Y-G. CRISPR/Cas9 platforms for genome editing in plants: Developments and applications. *Mol Plant*, 2016; 9(7): 961–974.
15. Char SN, Neelakandan AK, Nahampun H, Frame B, Main M, Spalding MH, et al. An Agrobacterium-delivered CRISPR/Cas9 system high-frequency targeted mutagenesis in maize. *Plant Biotechnol J.*, 2017; 15(2): 257–268.
16. Xie K, Minkenberg B, Yang Y. Boosting CRISPR/Cas9 multiplex editing capability with the endogenous tRNA-processing system. *Proc Natl Acad Sci USA.*, 2015; 112(11): 3570–3575.
17. Lowder LG, Zhang D, Baltes NJ, Paul JW 3rd, Tang X, Zheng X, et al. A CRISPR/Cas9 toolbox for multiplexed plant genome editing and transcriptional regulation. *Plant Physiol*, 2015; 169(2): 971–985.
18. Miao J, Guo D, Zhang J, Huang Q, Qin G, Zhang X, et al. Targeted mutagenesis in rice using a CRISPR-Cas system. *Cell Res.*, 2013; 23(10): 1233–1236.
19. Ormond KE, Mortlock DP, Scholes DT, Bombard Y, Brody LC, Faucett WA, et al. Human germline genome editing. *Am J Hum Genet.* 2017; 101(2): 167–176.
20. Shinagawa H, Makino K, Nakata A, Brenner S. Regulation of the Pho regulon in Escherichia coli K-12. *J Mol Biol.*, 1983; 168(3): 477–488.
21. Sancar A, Kacinski BM, Mott DL, Rupp WD. Identification of the *uvrC* gene product. *Proc Natl Acad Sci USA.*, 1981; 78(9): 5450–5454.
22. Ishino Y, Krupovic M, Forterre P. History of CRISPR-Cas from encounter with a mysterious repeated sequence to genome editing technology. *J Bacteriol* [Internet]. 2018 [cited 2020 Nov 2]; 200(7). Available from: <https://jb.asm.org/content/200/7/e00580-17>.
23. Barrangou R, Fremaux C, Deveau H, Richards M, Boyaval P, Moineau S, et al. CRISPR provides acquired resistance against viruses in prokaryotes. *Science*, 2007; 315(5819): 1709–1712.
24. Cong L, Zhang F. Genome engineering using CRISPR-Cas9 system. *Methods Mol Biol.*, 2015; 1239: 197–217.
25. Nitta M, Goto M, Shibuya N, Okawa Y. A novel protein with alkaline phosphatase and protease inhibitor activities in *Streptomyces hirosimensis*. *Biol Pharm Bull.*, 2002; 25(7): 833–836.
26. Lazdunski A. Peptidases and proteases of *Escherichia coli* and *Salmonella typhimurium*. *FEMS Microbiol Lett.*, 1989; 63(3): 265–276.
27. Dassa J, Fsihi H, Marck C, Dion M, Kieffer-Bontemps M, Boquet PL. A new oxygen-regulated operon in *Escherichia coli* comprises the genes for a putative third cytochrome oxidase and for pH 2.5 acid phosphatase (*appA*). *Mol Gen Genet.* 1991; 229(3): 341–352.
28. Piggot PJ, Sklar MD, Gorini L. Ribosomal alterations controlling alkaline phosphatase isozymes in *Escherichia coli*. *J Bacteriol.* 1972; 110(1): 291–299.
29. Messing SAJ, Ton-Hoang B, Hickman AB, McCubbin AJ, Peaslee GF, Ghirlando R, et al. The processing of repetitive extragenic palindromes: the structure of a repetitive extragenic palindrome bound to its associated nuclease. *Nucleic Acids Res.*, 2012; 40(19): 9964–9979.
30. Gupta RM, Musunuru K. Expanding the genetic editing tool kit: ZFNs, TALENs, and CRISPR-Cas9. *J Clin Invest.*, 2014; 124(10): 4154–4161.
31. Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A. Global cancer statistics, 2012: Global Cancer Statistics, 2012. *CA Cancer J Clin.* 2015; 65(2): 87–108.
32. Shine J, Dalgarno L. The 3'-terminal sequence of *Escherichia coli* 16S ribosomal RNA: complementarity to nonsense triplets and ribosome binding sites. *Proc Natl Acad Sci USA.*, 1974; 71(4): 1342–1346.
33. Wood EJ. Molecular cloning. A laboratory manual. *Biochem Educ.* 1983; 11(2): 82.
34. Hsu PD, Lander ES, Zhang F. Development and applications of CRISPR-Cas9 for genome engineering. *Cell.*, 2014; 157(6): 1262–1278.
35. Deveau H, Barrangou R, Garneau JE, Labonté J, Fremaux C, Boyaval P, et al. Phage response to CRISPR-encoded resistance in *Streptococcus thermophilus*. *J Bacteriol.* 2008; 190(4): 1390–400.
36. Deltcheva E, Chylinski K, Sharma CM, Gonzales K, Chao Y, Pirzada ZA, et al. CRISPR RNA maturation by trans-encoded small RNA and host factor RNase III. *Nature.* 2011; 471(7340): 602–607.
37. Gasiunas G, Barrangou R, Horvath P, Siksnys V. Cas9-crRNA ribonucleoprotein complex mediates specific DNA cleavage for adaptive immunity in bacteria. *Proc Natl Acad Sci USA.*, 2012; 109(39): E2579–2586.
38. DOE/Lawrence Berkeley National Laboratory. Programmable DNA scissors found for bacterial immune system. Science Daily [Internet]. 2012 Jun 28. [cited 2020 Nov 2]; Available from: <https://www.sciencedaily.com/releases/2012/06/120628193020.htm>.
39. Komor AC, Kim YB, Packer MS, Zuris JA, Liu DR. Programmable editing of a target base in genomic DNA without double-stranded DNA cleavage. *Nature.* 2016; 533(7603): 420–424.
40. Miller JH, editor. Experiments in Molecular Genetics. New York, NY: Cold Spring Harbor Laboratory Press, 1972.
41. Makino K, Shinagawa H, Amemura M, Nakata A. Nucleotide sequence of the *phoB* gene, the positive regulatory gene for the phosphate regulon of *Escherichia coli* K-12. *J Mol Biol.*, 1986; 190(1): 37–44.
42. Hart T, Tong AHY, Chan K, Van Leeuwen J, Seetharaman A, Aregger M, et al. Evaluation and design of genome-wide CRISPR/SpCas9 knockout screens. *G3 (Bethesda)*, 2017; 7(8): 2719–2727.

43. Doench JG, Fusi N, Sullender M, Hegde M, Vaimberg EW, Donovan KF, et al. Optimised sgRNA design to maximise activity and minimise off-target effects of CRISPR-Cas9. *Nat Biotechnol*, 2016; 34(2): 184–191.
44. Bäumllein H, Pustell J, Wobus U, Case ST, Kafatos FC. The 3' ends of two genes in the Balbiani ring c locus of *Chironomus thummi*. *J Mol Evol.*, 1986; 24(1–2): 72–82.
45. Meier JA, Zhang F, Sanjana NE. GUIDES: sgRNA design for loss-of-function screens. *Nat Methods*. 2017; 14(9): 831–832.
46. Heigwer F, Zhan T, Breinig M, Winter J, Brügemann D, Leible S, et al. CRISPR library designer (CLD): software for multispecies design of single guide RNA libraries. *Genome Biol.*, 2016; 17(1): 55.
47. Tao Y, Mis M, Blazer L, Ustav M Jnr, Steinhart Z, Chidiac R, et al. Tailored tetravalent antibodies potently and specifically activate Wnt/Frizzled pathways in cells, organoids and mice. *Elife* [Internet], 2019; 8. Available from: <http://dx.doi.org/10.7554/eLife.46134>
48. Sanjana NE, Wright J, Zheng K, Shalem O, Fontanillas P, Joung J, et al. High-resolution interrogation of functional elements in the non-coding genome. *Science*, 2016; 353(6307): 1545–1549.
49. Korkmaz G, Lopes R, Ugalde AP, Nevedomskaya E, Han R, Myacheva K, et al. Functional genetic screens for enhancer elements in the human genome using CRISPR-Cas9. *Nat Biotechnol*, 2016; 34(2): 192–198.
50. Europepmc.org. [cited Nov 2]. Available from: <https://europepmc.org/article/med/28976505>, 2020.
51. Ahn CH, Ramya M, An HR, Park PM, Kim Y-J, Lee SY, et al. Progress and challenges in the improvement of ornamental plants by genome editing. *Plants*, 2020; 9(6): 687.
52. Komor AC, Badran AH, Liu DR. CRISPR-based technologies for the manipulation of eukaryotic genomes. *Cell.*, 2017; 169(3): 559.
53. Shah F, Khan Z, Iqbal A, Turan M, Olgun M. Recent advances in grain crops research. Shah F, Khan Z, Iqbal A, Turan M, Olgun M, editors. *Intech Open*, 2020.
54. Yin H, Song C-Q, Dorkin JR, Zhu LJ, Li Y, Wu Q, et al. Therapeutic genome editing by combined viral and non-viral delivery of CRISPR system components in vivo. *Nat Biotechnol*, 2016; 34(3): 328–333.
55. Li B, Luo X, Deng B, Wang J, McComb DW, Shi Y, et al. An orthogonal array optimisation of lipid-like nanoparticles for mRNA delivery in vivo. *Nano Lett.*, 2015; 15(12): 8099–8107.
56. Liu J, Chang J, Jiang Y, Meng X, Sun T, Mao L, et al. Fast and efficient CRISPR/Cas9 genome editing in vivo enabled by bioreducible lipid and messenger RNA nanoparticles. *Adv Mater*, 2019; 31(33): e1902575.
57. Lander ES. The heroes of CRISPR. *Cell.*, 2016; 164(1–2): 18–28.
58. Ferreira R, David F, Nielsen J. Advancing biotechnology with CRISPR/Cas9: recent applications and patent landscape. *J Ind Microbiol Biotechnol*, 2018; 45(7): 467–480.
59. Liu H, Wang L, Luo Y. Blossom of CRISPR technologies and applications in disease treatment. *Synth Syst Biotechnol*, 2018; 3(4): 217–228.
60. BozorgQomi S, Asghari A, Mojarrad M. An overview of the CRISPR-based genomic- and epigenome-editing system: Function, applications, and challenges. *Adv Biomed Res.*, 2019; 8(1): 49.
61. McCarty NS, Graham AE, Studená L, Ledesma-Amaro R. Multiplexed CRISPR technologies for gene editing and transcriptional regulation. *Nat Commun*, 2020; 11(1): 1281.
62. Ethics and research integrity [Internet]. Ethics-and-integrity.org. [cited Nov 3]. Available from: <http://ethics-and-integrity.org/ethics/ethicsGenomeEditing.html>, 2020.
63. Li J-F, Norville JE, Aach J, McCormack M, Zhang D, Bush J, et al. Multiplex and homologous recombination-mediated genome editing in *Arabidopsis* and *Nicotianabenthiana* using guide RNA and Cas9. *Nat Biotechnol*, 2013; 31(8): 688–691.
64. Jiang W, Zhou H, Bi H, Fromm M, Yang B, Weeks DP. Demonstration of CRISPR/Cas9/sgRNA-mediated targeted gene modification in *Arabidopsis*, tobacco, sorghum and rice. *Nucleic Acids Res.*, 2013; 41(20): e188.
65. Fauser F, Schiml S, Puchta H. Both CRISPR/Cas-based nucleases and nickases can be used efficiently for genome engineering in *Arabidopsis thaliana*. *Plant J.*, 2014; 79(2): 348–359.
66. Song G, Jia M, Chen K, Kong X, Khattak B, Xie C, et al. CRISPR/Cas9: A powerful tool for crop genome editing. *Crop J.*, 2016; 4(2): 75–82.
67. Hyun Y, Kim J, Cho SW, Choi Y, Kim J-S, Coupland G. Site-directed mutagenesis in *Arabidopsis thaliana* using dividing tissue-targeted RGEN of the CRISPR/Cas system to generate heritable null alleles. *Planta*, 2015; 241(1): 271–284.
68. Feng Z, Mao Y, Xu N, Zhang B, Wei P, Yang D-L, et al. Multigeneration analysis reveals the inheritance, specificity, and patterns of CRISPR/Cas-induced gene modifications in *Arabidopsis*. *Proc Natl Acad Sci USA.*, 2014; 111(12): 4632–4637.
69. Mao Y, Zhang H, Xu N, Zhang B, Gou F, Zhu J-K. Application of the CRISPR-Cas system for efficient genome engineering in plants. *Mol Plant.*, 2013; 6(6): 2008–2011.
70. Kumar R, Kaur A, Pandey A, Mamrutha HM, Singh GP. CRISPR-based genome editing in wheat: a comprehensive review and future prospects. *Mol Biol Rep.*, 2019; 46(3): 3557–3569.

71. Gao J, Wang G, Ma S, Xie X, Wu X, Zhang X, et al. CRISPR/Cas9-mediated targeted mutagenesis in *Nicotianatabacum*. *Plant Mol Biol.*, 2015; 87(1–2): 99–110.
72. Sun X, Hu Z, Chen R, Jiang Q, Song G, Zhang H, et al. Targeted mutagenesis in soybean using the CRISPR-Cas9 system. *Sci Rep.*, 2015; 5(1): 10342.
73. Brooks C, Nekrasov V, Lippman ZB, Van Eck J. Efficient gene editing in tomato in the first generation using the clustered regularly interspaced short palindromic repeats/CRISPR-associated9 system. *Plant Physiol*, 2014; 166(3): 1292–1297.
74. Tabebordbar M, Zhu K, Cheng JKW, Chew WL, Widrick JJ, Yan WX, et al. In vivo gene editing in dystrophic mouse muscle and muscle stem cells. *Science*, 2016; 351(6271): 407–411.
75. Pickar-Oliver A, Gersbach CA. The next generation of CRISPR-Cas technologies and applications. *Nat Rev Mol Cell Biol.*, 2019; 20(8): 490–507.
76. Wirth T, Parker N, Ylä-Herttuala S. History of gene therapy. *Gene*, 2013; 525(2): 162–169.
77. Yu L, Tian X, Gao C, Wu P, Wang L, Feng B, et al. Genome editing for the treatment of tumorigenic viral infections and virus-related carcinomas. *Front Med*, 2018; 12(5): 497–508.
78. Cornu TI, Mussolino C, Cathomen T. Refining strategies to translate genome editing to the clinic. *Nat Med*, 2017; 23(4): 415–423.